

Bacterial Lipopolysaccharides and Neuron Toxicity in Neurodegenerative Diseases

Ian James Martins^{1,2,3*}

¹Centre of Excellence in Alzheimer's Disease Research and Care, Sarich Neuroscience Research Institute, Edith Cowan University, Verdun Street, Western Australia, Australia.

²School of Psychiatry and Clinical Neurosciences, The University of Western Australia, Nedlands, Australia.

³McCusker Alzheimer's Research Foundation, Hollywood Medical Centre, 85 Monash Avenue, Nedlands, 6009, Australia.

***Correspondence:**

Dr. Ian Martins, School of Medical Sciences, Edith Cowan University, Western Australia 6009, Australia, Tel: +61863042574, E-mail: i.martins@ecu.edu.au.

Received: 09 June 2018; **Accepted:** 05 July 2018

Citation: Ian James Martins. Bacterial Lipopolysaccharides and Neuron Toxicity in Neurodegenerative Diseases. *Neurol Res Surg.* 2018; 1(1): 1-3.

Keywords

Neuron death, Bacterial lipopolysaccharides, Sirtuin 1, Protein aggregation, Mitophagy.

Editorial

Research into the role of bacterial lipopolysaccharides (LPS) has escalated with the improved understanding of the LPS effects on astrocytes and neurons with relevance to brain neuroinflammation [1,2] that is now of major concern in neurodegenerative diseases. LPS activates the toll-like receptor 4 (TLR-4) in mouse and human astrocytes with TLR-4 linked to neuroinflammation and neuron death [3-6]. LPS related toxicity may influence neuron membrane cholesterol by binding to cell membranes that promote amyloid beta (Aβ) aggregation and fibril formation [7] to mediate accelerated neuron death. LPS are endotoxins and essential components of the outer membrane of gram negative bacteria and consist of covalently linked segments, surface carbohydrate polymer, core oligosaccharide and acylated glycolipid that can bind to cell membranes to alter membrane interactions [8,9]. LPS regulate plasma acute phase proteins (gelsolin, serum amyloid protein A, serum amyloid protein, C-reactive protein, clusterin, transthyretin) [7] and various other acute phase proteins (APP) involved in Aβ aggregation (transferrin, albumin, phospholipid transfer protein, LPS binding protein (LBP), albumin).

The cluster of differentiation 14 (CD14) receptor is referred to as the LPS receptor and involved with brain Aβ metabolism [7]. The CD14 receptor assists in the co-ordination of the microglia that promotes Aβ mediated and oxidative neuron death [10]. In the developing world increased plasma LPS levels have raised major concern with relevance to CD14 regulation of TLR-4 mediated accelerated neuron death [11,12]. Detailed studies

now indicate that LPS now repress the nuclear receptor Sirtuin 1 (Sirt 1) with its toxic effects related to interference with Sirt 1's role in the regulation of transcription factors [13] related to neuron proliferation and induction of Type 3 diabetes. Sirt 1 is now closely linked to the immune system [14], insulin resistance and metabolic activity [7]. Sirt 1's role in neuron death is now connected to cellular proteins (Figure 1) such as heat shock protein (HSP), cellular prion protein (PrPc), alpha-synuclein and tau that are connected to Aβ aggregation [15-23] and accelerated neuron death. LPS represses Sirt 1 [13] with HSP involved in the regulation of PrPc and Aβ aggregation relevant to mitochondrial apoptosis and neuron death [24-39]. The nuclear receptor Sirt 1's role on neuron survival/apoptosis via LPS is primary with effects of LPS secondary on CD14 regulation of TLR-4 mediated neuron apoptosis [7,10-12].

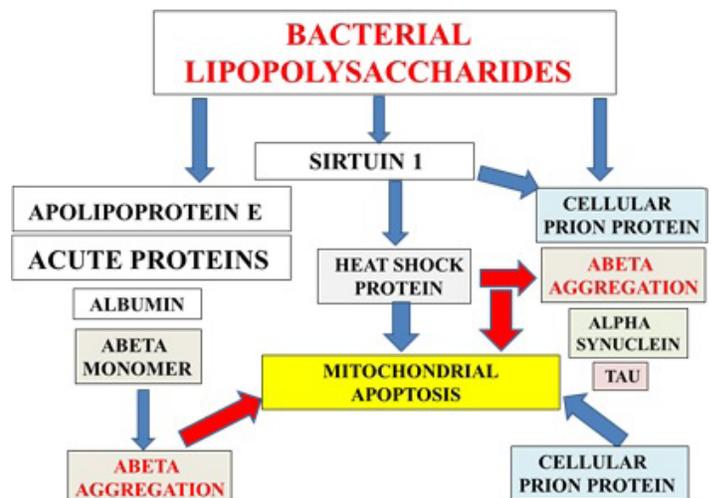


Figure 1: LPS neutralize apolipoprotein E and repress the nuclear receptor Sirt 1 relevant to protein aggregation and mitochondrial apoptosis. The effects of LPS on mitophagy may be mediated by acute phase proteins, albumin, cellular prion protein or amyloid beta aggregation. Under core body temperature disturbances heat shock proteins induce amyloid beta/cellular prion protein aggregation and induce neuronal mitophagy with relevance to neurodegenerative disease and epilepsy induced stroke.

LPS and its involvement with Sirt 1 in protein aggregation is linked to magnesium deficiency [40] with induction of mitochondrial apoptosis associated with epilepsy induced stroke [41-44]. Other Sirt 1 inhibitors such as palmitic acid, suramin, sirtinol, alcohol and fructose should be carefully controlled to prevent mitochondrial apoptosis [45] and neuron death. Excessive consumption of arginine, patulin, xenobiotics and butyric acid should be avoided to prevent Sirt 1 downregulation. Excessive caffeine consumption should be avoided with relevance to magnesium deficiency [45] and nuclear receptor Sirt 1 disturbances associated with Type 3 diabetes [45]. The effects of core body temperature inactivate Sirt 1 with HSP and PrPc connected to protein aggregation [15,46-49] and mitophagy in neurodegenerative diseases and epilepsy induced stroke [42].

Conclusion

Toxic protein aggregation and neuron death has become of major concern to neurological stroke, cerebrovascular diseases, neurological disorders and functional/epilepsy Res Surgery. Plasma LPS should be monitored early in life to prevent mitochondrial apoptosis in neurodegenerative diseases. Dietary and pharmacological inhibition of neuron nuclear receptors will determine neuron proliferation and remodeling with excessive nuclear receptor inhibitor consumption related to protein aggregation in neurological diseases. Core body temperature malfunction will lead to uncontrolled aggregation of proteins with neuronal mitophagy associated with neurological stroke.

Acknowledgments

This work was supported by grants from Edith Cowan University, the McCusker Alzheimer's Research Foundation and the National Health and Medical Research Council.

References

1. Murray CL, Skelly DT, Cunningham C. Exacerbation of CNS inflammation and neurodegeneration by systemic LPS treatment is independent of circulating IL-1 β and IL-6. *J Neuroinflammation*. 2011; 8: 50.
2. Qin L, Wu X, Block ML, et al. Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. *Glia*. 2007; 55: 453-462.
3. Vasconcelos AR, Yshii LM, Viel TA, et al. Intermittent fasting attenuates lipopolysaccharide-induced neuroinflammation and memory impairment. *Journal of Neuroinflammation*. 2014; 11: 85.
4. Tanaka S, Ide M, Shibutani T, et al. Lipopolysaccharide-induced microglial activation induces learning and memory deficits without neuronal cell death in rats. *J Neurosci Res*. 2006; 83: 557-566.
5. Walter S, Letiembre M, Liu Y, et al. Role of the toll-like receptor 4 in neuroinflammation in Alzheimer's disease. *Biochem*. 2007; 20: 947-956.
6. Gorina R, Font-Nieves M, Márquez-Kisinousky L, et al. Astrocyte TLR4 activation induces a proinflammatory environment through the interplay between MyD88-dependent NF κ B signaling, MAPK, and Jak1/Stat1 pathways. *Glia*. 2011; 59: 242-255.
7. Martins IJ. Unhealthy Diets Determine Benign or Toxic Amyloid Beta States and Promote Brain Amyloid Beta Aggregation. *Austin J Clin Neurol*. 2015; 2: 1060-1066.
8. Martins IJ. LPS regulates apolipoprotein E and amyloid beta interactions with effects on acute phase proteins and amyloidosis. *AAR*. 2015; 4: 69-77.
9. Martins IJ. Bacterial Lipopolysaccharides Change Membrane Fluidity with Relevance to Phospholipid and Amyloid Beta Dynamics in Alzheimer's Disease. *J Microb Biochem Technol*. 2016; 8: 322-324.
10. Bate C, Veerhuis R, Eikelenboom P, et al. Microglia kill amyloid-beta1-42 damaged neurons by a CD14-dependent process. *Neuroreport*. 2004; 15: 1427-1430.
11. Zanoni I, Ostuni R, Lorri R, et al. CD14 controls the LPS-induced endocytosis of Toll-like Receptor 4. *Cell*. 2011; 147: 868-880.
12. Arroyo-Espliguero R, Avanzas P, Jeffery S, et al. CD14 and toll-like receptor 4: a link between infection and acute coronary events? *Heart*. 2004; 90: 983-988.
13. Martins IJ. The Future of Genomic Medicine Involves the Maintenance of Sirtuin 1 in Global Populations. *Int J Mol Biol*. 2017; 2: 00013.
14. Martins IJ. Antimicrobial activity inactivation and toxic immune reactions induce Epilepsy in human. *J Med Discov*. 2017; 2: 1-7.
15. Martins IJ. Heat Shock Gene Inactivation and Protein Aggregation with Links to Chronic Diseases. *Diseases*. 2018; 6: 1-5.
16. Gasperini L, Legname G. Prion protein and aging. *Front Cell Dev Biol*. 2014; 2: 44.
17. Chen D, Steele AD, Hutter G, et al. The role of calorie restriction and SIRT1 in prion mediated neurodegeneration. *Exp Gerontol*. 2008; 43: 1086-1093.
18. Seo JS, Moon MH, Jeong JK, et al. SIRT1, a histone deacetylase, regulates prion protein-induced neuronal cell death. *Neurobiol Aging*. 2012; 33: 1110-1120.
19. Martins IJ. Diabetes and Cholesterol Dyshomeostasis Involve Abnormal α -Synuclein and Amyloid Beta Transport in Neurodegenerative Diseases. *Austin Alzheimers J Parkinsons Dis*. 2015; 2: 1020-1028.
20. Aulić S, Masperone L, Narkiewicz J, et al. α -Synuclein Amyloids Hijack Prion Protein to Gain Cell Entry, Facilitate Cell-to-Cell Spreading and Block Prion Replication. *Sci Rep*. 2017; 7: 1-12.
21. De Cecco E, Legname G. The role of the prion protein in the internalization of α -synuclein amyloids. *Prion*. 2018; 12: 23-27.

22. Urrea L, Ferrer I, Gavín R, et al. The cellular prion protein (PrPC) as neuronal receptor for α -synuclein. *Prion*. 2017; 11: 226-233.
23. Jellinger KA, Popescu BO. Interaction between pathogenic proteins in neurodegenerative disorders. *J Cell Mol Med*. 2012; 16: 1166-1183.
24. Salazar SV, Strittmatter SM. Cellular prion protein as a receptor for amyloid- β oligomers in Alzheimer's disease. *Biochem Biophys Res Commun*. 2017; 483: 1143-1147.
25. Purro SA, Nicoll AJ, Collinge J. Prion Protein as a Toxic Acceptor of Amyloid- β Oligomers. *Biol Psychiatry*. 2018; 83: 358-368.
26. Kessels HW, Nguyen LN, Nabavi S, et al. The prion protein as a receptor for amyloid-beta. *Nature*. 2010; 466: E3-4-E4-5.
27. Jarosz-Griffiths HH, Noble E, Rushworth JV, et al. Amyloid- β Receptors: The Good, the Bad, and the Prion Protein. *J Biol Chem*. 2015; 291: 3174-3183.
28. Falker C, Hartmann A, Guett I, et al. Exosomal cellular prion protein drives fibrillization of amyloid beta and counteracts amyloid beta-mediated neurotoxicity. *J Neurochem*. 2016; 137: 88-100.
29. Laurén J, Gimbel DA, Nygaard HB, et al. Cellular prion protein mediates impairment of synaptic plasticity by amyloid- β oligomers. *Nature* 2009; 457: 1128-1132.
30. Nicoll AJ, Silvia Panico S, Freir DB, et al. Amyloid- β nanotubes are associated with prion protein-dependent synaptotoxicity. *Nat Commun*. 2013; 4: 2416.
31. Saleem F, Bjorndahl TC, Ladner CL. Lipopolysaccharide induced conversion of recombinant prion protein. *Prion*. 2014; 8: 221-232.
32. Ladner-Keay CL, LeVatte M, Wishart DS. Role of polysaccharide and lipid in lipopolysaccharide induced prion protein conversion. *Prion*. 2016; 10: 466-483.
33. Chen, S, Yadav SP, Surewicz WK. Interaction between human prion protein and amyloid-beta (Abeta) oligomers: role OF N-terminal residues. *J Biol Chem*. 2010; 285: 26377-26383.
34. Younan ND, Sarell CJ, Davies P, et al. The cellular prion protein traps Alzheimer's Abeta in an oligomeric form and disassembles amyloid fibers. *FASEB J*. 2013; 27: 1847-1858.
35. Bove-Fenderso E, Urano R, Straub JE, et al. Cellular prion protein targets amyloid- β fibril ends 892 via its C-terminal domain to prevent elongation. *J Biol Chem*. 2017; 292: 16858-16871.
36. Faris R, Moore RA, Ward A, et al. Cellular prion protein is present in mitochondria of healthy mice. *Sci Rep*. 2017; 7: 41556.
37. Hachiya NS, Yamada M, Watanabe K, et al. Mitochondrial localization of cellular prion protein (PrPC) invokes neuronal apoptosis in aged transgenic mice overexpressing PrPC. *Neurosci Lett*. 2005; 374: 98-103.
38. De Mario A, Peggion C, Massimino ML, et al. The prion protein regulates glutamate-mediated Ca²⁺ entry and mitochondrial Ca²⁺ accumulation in neurons. *J Cell Sci*. 2017; 130: 2736-2746.
39. Bertoli A, Sorgato MC. Neuronal pathophysiology featuring PrPC and its control over Ca²⁺ metabolism. *Prion*. 2018; 12: 28-33.
40. Martins IJ. Magnesium Therapy Prevents Senescence with the Reversal of Diabetes and Alzheimer's Disease. *Health*. 2016; 8: 694-710.
41. Walz R, Castro RM, Velasco TR, et al. Cellular prion protein: implications in seizures and epilepsy. *Cell Mol Neurobiol*. 2002; 22: 249-257.
42. Martins IJ. Food Quality and Advances in Pharmacological Management Prevent Mitochondrial Apoptosis and Epilepsy Induced Stroke. *Research and Reveiws: Neuroscience*. 2018; 2: 7-9.
43. Martins IJ. Bacterial LPS Overrides Adenosine Treatment of Epileptic Seizures. *EC Microbiology*. 2017; 7: 83-86.
44. Martins IJ. Sirtuin 1 and Adenosine in Brain Disorder Therapy. *J Clin Epigenet*. 2017; 3:1-11.
45. Martins IJ. Nutrition Therapy Regulates Caffeine Metabolism with Relevance to NAFLD and Induction of Type 3 Diabetes. *J Diabetes Metab Disord*. 2017; 4: 019.
46. Kenward N, Landon M, Laszlo L, et al. Heat shock proteins, molecular chaperones and the prion encephalopathies. *Cell Stress Chaperones*. 1996; 1: 18-22.
47. Shyu WC, Harn HJ, Saeki K, et al. Molecular modulation of expression of prion protein by heat shock. *Mol Neurobiol*. 2002 ; 26: 1-12.
48. Chamachi NG, Chakrabarty S. Temperature-Induced Misfolding in Prion Protein: Evidence of Multiple Partially Disordered States Stabilized by Non-Native Hydrogen Bonds. *Biochemistry*. 2017; 56: 833-844.
49. Moulick R, Udgaonkar JB. Thermodynamic characterization of the unfolding of the prion protein. *Biophys J*. 2014; 106: 410-420.